

UB Next-Generation Sequencing and Expression Analysis Core Roche/454 GS FLX Titanium Sequencing Sample Submission Form

Please print both pages of this form and bring a completed and signed copy with you when you drop-off or ship samples to the UB Next-Gen Sequencing and Expression Analysis Core for GS FLX Titanium Sequencing Services. We will not accept samples unless accompanied by this signed and completed form.

Lab Contact Information

PI Name _____

Technician Name _____

Institution/Business _____

Phone _____

Address _____

Email _____

City/State/Zip _____

Phone _____

SUNY at Buffalo Researchers Only – How will you be paying for your GS FLX Titanium Sequencing Services (Research Foundation Account, State Account, etc.)?

Email _____

Experimental Design Information

What type of GS FLX Titanium Run will you be doing (circle one)?

Shotgun 3kb Paired End 8kb Paired End 20 kb Paired End Amplicon

How many sequencing runs? _____ How many samples? _____

Do you want to use MIDs (circle one)? Yes No

How many regions should the sequencing plate be divided into (circle one)? 2 4 6 8

Provide a brief description of your experimental design in the space below. Please include your multiplexing plans making sure to describe how you want to use MIDs and which samples should be pooled, if applicable. If more space is needed for the description, you may attach additional pages to this form.

Sample Information

The following information is required for each sample that you are bringing to the UB Next-Gen Sequencing and Expression Analysis Core for GS FLX Titanium Sequencing Services. For all sample types, confirm that each sample name is unique and that the name written on this form matches the name written on the sample tube. Provide the sample volume, concentration, and measurement method. Attach an agarose gel image of the samples to this form and make sure to clearly label the sample wells and ladder sizes. For shotgun and paired end samples only; provide the 260/280 ratio and identify the method that was used for DNA extraction or indicate that you would like the UB Next-Gen Sequencing Core to extract DNA from bacterial pellets provided by the researcher. If you used a method other than the recommended Qiagen Genomic Buffers Set in conjunction with the Qiagen Genomic-Tip Protocol, attach a copy of your protocol and confirm that your samples have been RNase treated. For amplicon samples only; attach a description of your PCR amplification conditions and list all of your primer sequences. Indicate which amplicons were amplified with which primers, the exact amplicon size (base pairs), and which samples should be pooled together. If you are unable to provide any of the required information for your samples you must contact us prior to dropping off samples to discuss alternative options.

